

## **Nanometric grafting of poly (N-isopropylacrylamide) onto polystyrene film by different doses of gamma radiation**

**G.R. Shahhosseini<sup>1,\*</sup>, F. Asadifard<sup>2</sup>, M. Enayatollahi<sup>3</sup>, M. Shapourgan<sup>3</sup>,  
R. Haghjo<sup>3</sup>, S. Heidari Keshel<sup>4</sup>**

<sup>1</sup>Agricultural , Medical And Industrial Research School – Nuclear Science And Techniques Research Institute–Karaj, Iran

<sup>2</sup>Nursing and Midwifery Faculty, Karaj Islamic Azad University, Karaj, Iran

<sup>3</sup>Biomedical Engineering Faculty, Islamic Azad University Science and Research Branch, Tehran, Iran

<sup>4</sup>Stem cell preparation unit, Farabi Hospital, Tehran University Medical Science, Tehran, Iran

Received: 4 April 2010; Accepted: 28 July 2010

---

### **Abstract**

Poly N-isopropylacrylamide was successfully grafted onto a polystyrene cell culture dish by gamma ray. In this study, the effect of a radiation dose (radiation absorbed dosages of 10, 20, 30, 40 KGy) under appropriate temperature and grafting conditions was investigated. The FTIR analysis showed the existence of the graft PNIPAAm on the substrate. The optimal value of the dose for grafting was 40 KGy at 50°C. The SEM and AFM images clearly showed that increasing the absorbed dose of radiation would increase the amount of grafting. Surface topography and graft thickness in AFM images of the radiated samples showed that the PNIPAAm at the absorbed dose of radiation was properly grafted. The thickness of these grafts was about 50–100 nm. The drop water contact angles of the best grafted sample at 37°C and 10°C were  $55.3 \pm 1.2^\circ$  and  $61.2 \pm 0.9^\circ$  respectively, which showed the hydrophilicity and hydrophobicity of the grafted surfaces. DSC analysis also revealed the LCST of the grafted sample to be 32°C. Thermo-responsive polymers were grafted to dishes covalently which allowed fibroblast cells to attach and proliferate at 37°C; the cells also detached spontaneously without using enzymes when the temperature dropped below 32°C. This characteristic proves that this type of grafted material has potential as a biomaterial for cell sheet engineering.

**Keywords:** *Nanometric grafting, PNIPAAm, polystyrene film, gamma ray, cell engineering*

---

### **1. Introduction**

During recent decades several materials and medical devices have been produced for medical purposes. Cell sheet engineering has been developed to avoid tissue reconstruction limitations using biodegradable scaffolds or single cell suspension injection [1-4]. Cell sheets are developed by thermo-responsive culture dishes. Thermo-responsive polymers are grafted to dishes

---

\* Corresponding author: Gholamreza Shahhosseini

Agricultural , Medical And Industrial Research

School – Nuclear Science And Techniques

Research Institute, Iran.

Tel +98 2614470802

Email [gshahosseini@yahoo.com](mailto:gshahosseini@yahoo.com)

covalently, which allows different cell types to attach and proliferate at 37°C. Cells detach spontaneously without using enzymes when the temperature decreases below 32°C; this is due to the natural specification of the intelligent polymers, and also to the detachment of the cell metabolic changes made by the polymer resulting from decreasing temperature [5-9]. Environmentally-sensitive systems or intelligent polymers are those that react to small environmental changes. Those functional polymers that react to their readjustment or physical and chemical changes in the environment are generally known as stimuli-responsive or intelligent polymers. Thermo-responsive polymers show a balanced and proper hydrophilic-hydrophobic in their structures. They are able to switch on-off the receptor using the transition between the extended and coiled forms of the molecule [10-12]. NIPAAm and its copolymers are among those materials which have LCST. PNIPAAm shows LCST at 32°C. While the temperature is over 32°C, the polymer is solid and hydrophobic but when it is below 32°C it is completely hydrated and shows hydrophilic properties [13]. Polymer grafting gives considerable thermo-responsive features to surfaces. One of the methods used to create intelligent surfaces is grafting monomers onto polymer surfaces such as PET and PP. Chemical and physical polymerization methods such as glow discharge [14], corona discharge [15], high energy [16], ozone [17] and UV [18,9] have been followed to graft monomers on surfaces. Another important method is radiation diffusion in the material which causes production of secondary electrons, which leads to molecule ionization. Usually on a radiated substrate, free radicals and peroxide groups are produced. During a deoxygenating process using N<sub>2</sub> pre-irradiated substrates are suspended in a monomer suspension for specific periods of time at the proper temperature [8]. PNIPAAm and polyacrylic acid are successfully grafted onto different substrates such as PTFE [19] and PVDF [20] or PET [21] using radiation. In this study, the monomer of NIPAAm has been grafted onto a polystyrene substrate using different absorbed doses of radiation.

## 2. Materials and methods

### 2.1. Materials

Polystyrene dishes with the dimensions of 1×1 cm<sup>2</sup> and 1 mm thickness, ethanol and methanol (Merck Co), NIPAAm (Aldrich co), n-hexane (Merck co), distilled water, polystyrene and fibroblast cells (L929) were used in this study. Polystyrene dishes were put in a solution of ethanol-methanol with a 50/50 ratio for 24 hours to dissolve impurities and oils existing on the surface of the dishes. After removal from the solution, the dishes were washed in distilled water. For recrystallization of NIPAAm, 10.3 g of NIPAAm (Aldrich Co) was dissolved in 125 mL n-hexane and the solution was then placed in a refrigerator to make the NIPAAm ready for grafting.

### 2.2. Irradiation

A <sup>60</sup>Co gamma radiation source, supplied by Karaj Atomic Research Centre, Iran, was used for the irradiation of the samples. The dose rate was 1 KGy/h. Irradiations were carried out in the air under ambient conditions.

### 2.3. Graft polymerization

In this study,  $^{60}\text{Co}$ - $\gamma$ -radiation with a radiating absorbed dose of 1 KGy/h was used. Pre-irradiated polystyrene samples (for 10, 20, 30, 40 hours) were put in a preprepared solution including recrystallised NIPAAm with distilled water which was degassed by nitrogen gas (2 bar mass flow rate) for 20 minutes. This process was done to increase the efficiency of the free radical polymerization (deoxygenation). The samples in this solution were put in a water bath at  $50^{\circ}\text{C}$  for 2 hours; then the samples were brought out, washed in distilled water and put into distilled water for 72 hours, after which they were taken out for analysis. The effect of the  $\gamma$ -radiation dose on the grafting degree was calculated using the following formula:

Grafting (%) =  $(w-w^{\circ}) 100/w^{\circ}$ , where  $w$  and  $w^{\circ}$  indicate the weights of the grafted and ungrafted samples respectively.

### 2.4. Fourier transmission infrared spectroscopy (FTIR)

The samples were examined by FTIR (Bruker-Equinox 55) before and after adjustment. The samples were scratched into powder and were produced as capsule using KBr, and then, were put to investigation.

### 2.5. Scanning electron microscopy (SEM) and Atomic force microscopy (AFM)

The surface characteristics of various modified and unmodified films were studied with the help of SEM (Cambridge Stereo scan, model S-360) to analyze the changes in the surface morphology. The films were first coated with a gold layer (Joel fine coat, ion sputter for 2 hours) to provide surface conduction before their scanning. The surface topology characteristics and the thickness of various modified films as well as the unmodified films were studied with the help of AFM (TMX 2010) to analyze the changes in the surface topology.

### 2.6. Contact angle analysis

The sample surfaces' static contact angles were investigated by a contact angle measuring device (Kruss G10) following the sessile drop method. The formed contact angle was defined as the angle between solid/liquid and liquid/vapor join surface. In order to review the sample surfaces' hydrophilic/hydrophobic behavior at high and low temperatures, the samples were examined at two different temperatures of  $10^{\circ}\text{C}$  and  $37^{\circ}\text{C}$  and the contact angles were measured at these temperatures.

### 2.7. Differential scanning calorimetry (DSC)

The samples were investigated by thermal analysis using a DSC device (Netzschdsc200F3) with a heating rate of 5 degree per minute from  $0^{\circ}\text{C}$  to  $60^{\circ}\text{C}$  in a nitrogen gas atmosphere.

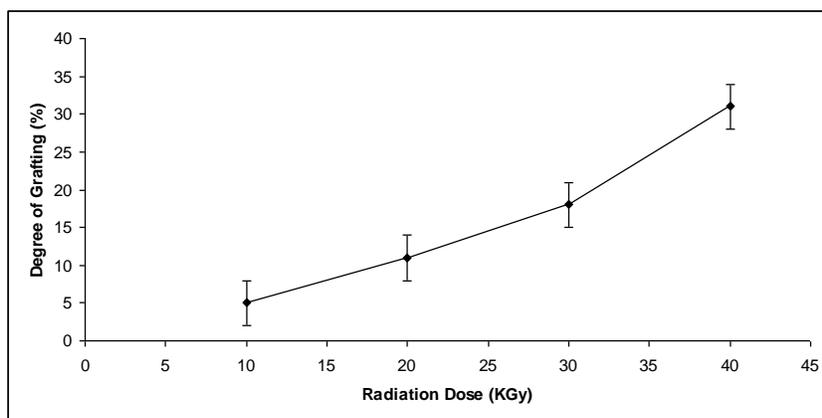
### 2.8. Cell investigations

The fibroblast cell suspension (L929) from a mouse tail was prepared according to the ISO10993 standard. The fibroblast cell suspension was transferred to a flask (25 cc) containing 5 mL DMEM (2Mm l-glutamine, penicillin (100 lu/mL), streptavidin (100 $\mu\text{L}$ /mL)) and FBS 10%. The suspension was then placed in an incubator (5%  $\text{CO}_2$ ,  $37^{\circ}\text{C}$ ). The fibroblast cells were proliferated in the flask and were washed using FBS/EDTA. Then the trypsin enzyme/EDTA was added to the flask ( $4^{\circ}\text{C}$ ), and the flask was incubated for 2 min. The culture media (FBS/DMEM)

was added to the flask, and the cells were gently pipetted. The cell suspension was transferred to a falcon tube (15 mL) and centrifuged (1410 rpm) for 5 min. The solution was removed and the precipitation was transferred to a new flask (75 cc) for reculturing. The surface of the samples was well cleaned using cotton and alcohol. Pieces of cell culture (0.5×0.5 cm) from the petri dish (control) and the main sample were cut and placed individually in one of the Petri dish wells by using a sterilized pincer. 50000 cells/well were seeded into a 12-well culture plate, removed by pipette and were poured onto the control and the main samples. Then all of the samples were placed in a Memmert incubator at 37°C for 48 hours. The samples were removed from the incubator and refrigerated at 10°C for 2 hours before being studied through a Nikon Eclipse Ts-100 photonic microscope.

### 3. Results and discussion

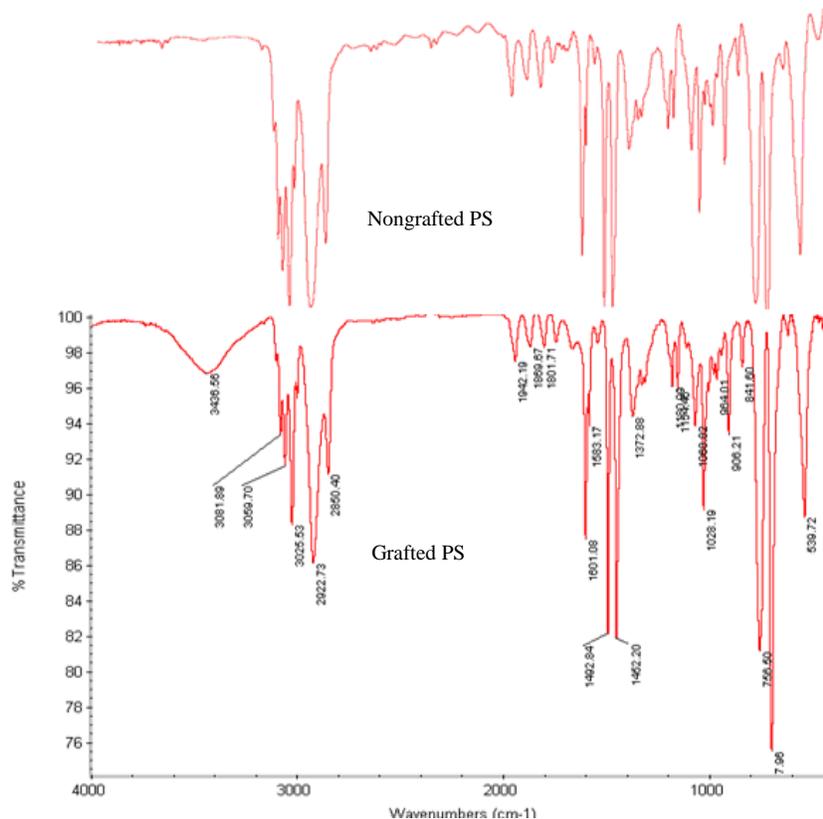
The pre-irradiated polystyrene samples dosed at 10, 20, 30 and 40 KGy were weighed at a constant temperature. **Figure 1** shows the grafting increase rate versus the increase in radiation dosage. The maximum rate of graft growth on the polystyrene substrate (average 31%) was achieved under the highest dosage of radiation (40 KGy).



**Fig.1.** Grafting as a function of dose, at a reaction temperature of 50°C

#### 3.1. Fourier transmission infrared spectroscopy (FTIR)

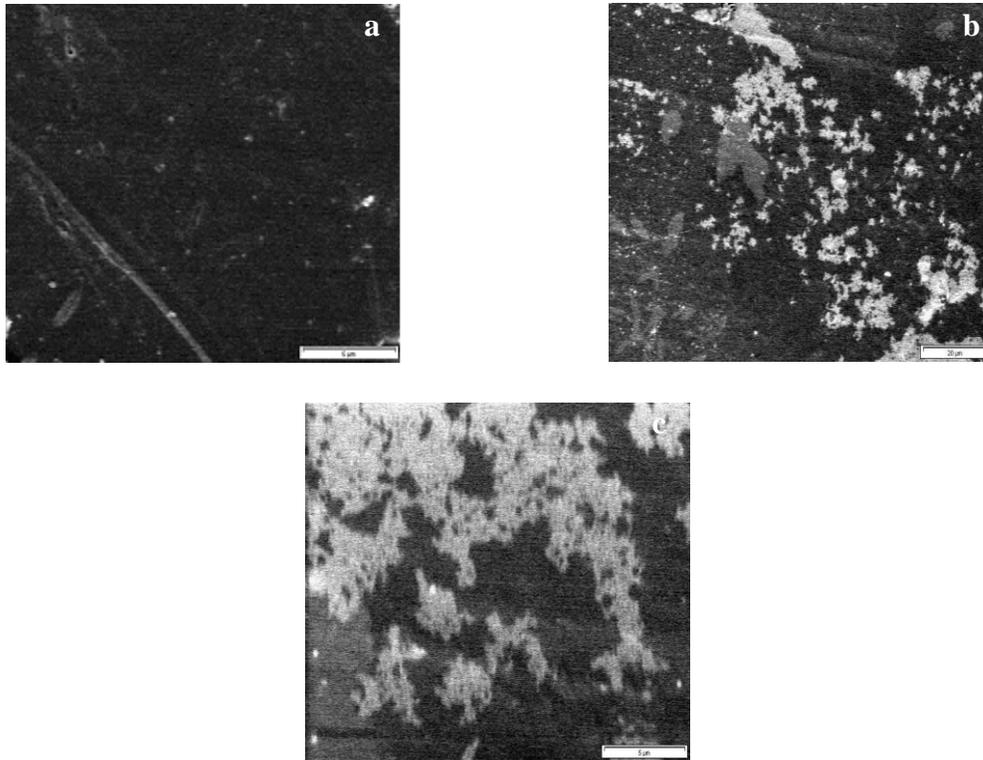
FTIR spectra results of the regular unadjusted and the  $\gamma$ -radiation-adjusted polystyrene samples are shown in **Figure 2**. The FTIR spectra of the NIPAAm grafted by the  $\gamma$ -radiated polystyrene are shown in the lower part of **Figure 2**. The PNIPAAm characteristic points include 1601  $\text{cm}^{-1}$  which indicate  $-\text{NH}$  groups, 1730-1830  $\text{cm}^{-1}$  which indicate  $\text{C}=\text{O}$  groups, 3025  $\text{cm}^{-1}$  which indicate  $\text{CH}_3$  groups, and 3443  $\text{cm}^{-1}$  which indicate  $\text{NH}$  groups, in PNIPAAm. All these points are found in the PNIPAAm-grafted polystyrene samples. This demonstrates grafting between PNIPAAm and the polystyrene surface through  $\gamma$ -radiation coating activation.



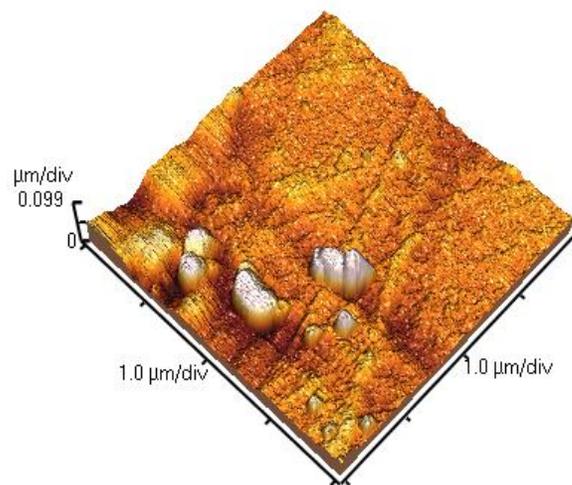
**Fig. 2.** FTIR spectra of the nongrafted polystyrene (upper) and grafted polystyrene treated with gamma rays (radiation dose: 40 KGy) (lower)

### 3.2. Scanning electron microscopy (SEM) and atomic force microscopy (AFM)

The images for investigating the adjusted samples through  $\gamma$ -radiation are shown in [Figure 3](#), which shows the PNIPAAm graft at different radiation dosages in polystyrene. [Figure 3a](#) is the SEM image obtained from a normal polystyrene substrate sample; the visible lines indicate slight superficial scratches which are clearly seen in a 5000x magnification. The SEM images of the PNIPAAm-grafted samples show the existence of the grafted PNIPAAm on the polystyrene surfaces. The quantity of the radiated-absorbed dose of 10 KGy is very low, as is clearly shown in [Figures 3b](#) and [c](#) by the number of white spots in the 1000x and 5000x magnifications of the SEM images of the grafted surfaces. The surfaces' topography and the created graft thickness on the surface as shown in the AFM images ([Figure 4](#)) also confirms this statement. The average graft thickness was about 100 nm and the white spots indicate the roughness created during radiation.

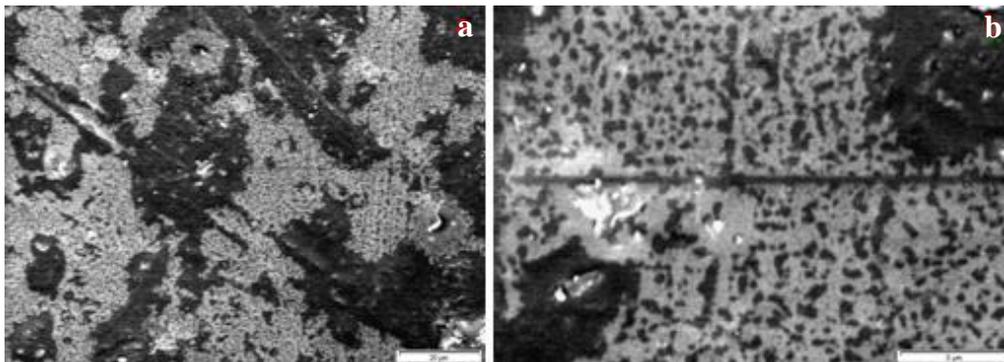


**Fig.3.** **a.** Scanning electron microscopy of non grafted polystyrene. Magnification 5000x. **b.** Scanning electron microscopy of grafted polystyrene under 10 KGy. Magnification 1000x (scale: 20 μm), **c.** Magnification 5000x (scale: 5 μm)

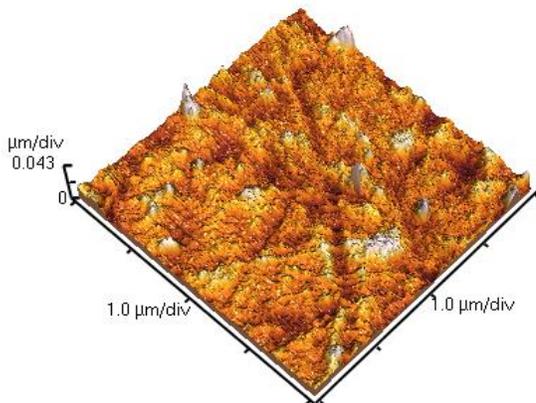


**Fig.4.** Atomic force microscopy of grafted polystyrene under 10 KGy. (scale: 1×1 μm)

Figures 5 and 6 show the SEM and the AFM images of the PNIPAAm-grafted samples under a radiation-absorbed dose of 20 KGy. These images demonstrate that the graft quantity was increased, as is clearly visible in the 1000x and 5000x magnifications. The topography of the surfaces shown in the AFM images indicates roughness created as a result of PNIPAAm grafting on the polystyrene surfaces. The observed graft thickness in these AFM images is about 45 nm.

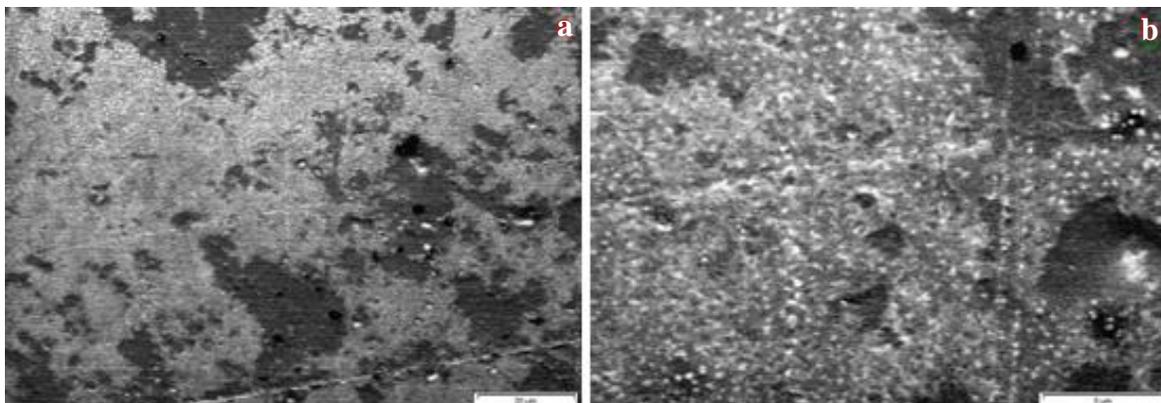


**Fig.5.** Scanning electron microscopy of grafted polystyrene under 20 KGy, **a.** Magnification 1000x (scale: 20  $\mu\text{m}$ ), **b.** Magnification 5000x (scale:5  $\mu\text{m}$ )

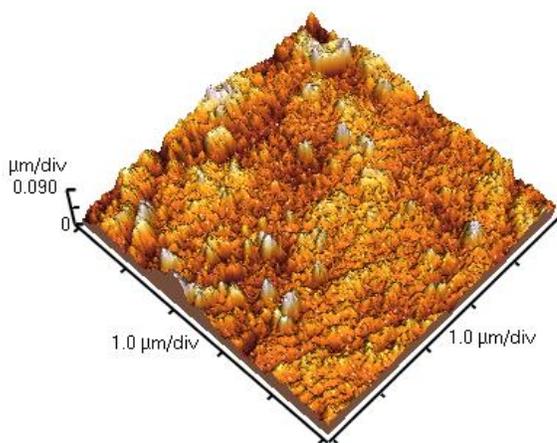


**Fig.6.** Atomic force microscopy of grafted polystyrene under 20 KGy. (scale: 1 $\times$ 1  $\mu\text{m}$ )

Figures 7 and 8 show images of SEM and AFM, PNIPAAm-grafted samples under a radiation absorbed dose of 30 KGy. According to the images, the quantity of the graft has been increased. The surface topography and the created thickness on the surface is shown in the AFM images. These also indicate more rough surfaces which could be the result of a complete PNIPAAm graft on the polystyrene surface. The observed graft thickness in the AFM images is about 90 nm.

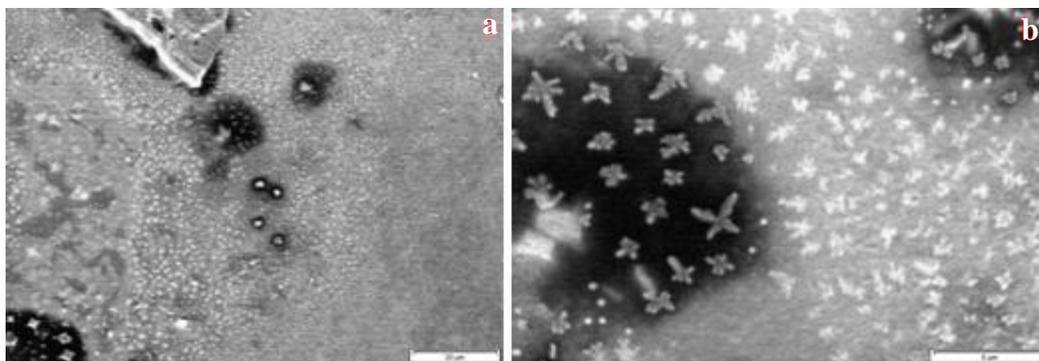


**Fig.7.** Scanning electron microscopy of grafted polystyrene under 30 KGy, **a.** Magnification 1000x (scale: 20  $\mu\text{m}$ ), **b.** Magnification 5000x (scale: 5  $\mu\text{m}$ )

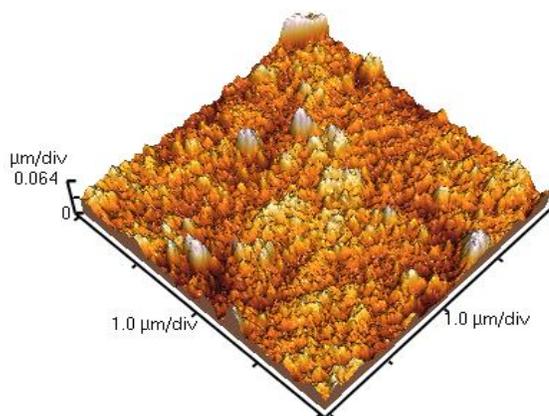


**Fig.8.** Atomic force microscopy of grafted polystyrene under 30 KGy. (scale:  $1 \times 1 \mu\text{m}$ )

Figures 9 and 10 show the SEM and AFM images of PNIPAAm-grafted samples under a radiation absorbed dose of 40 KGy. According to the images the quantity of the graft has been increased. The surface topography and the created thickness on the surface is shown in the AFM images. These also indicate more rough surfaces which could be the result of a complete PNIPAAm graft on the polystyrene surface. The observed graft thickness in the AFM images is about 60 nm.



**Fig.9.** Scanning electron microscopy of grafted polystyrene under 40 KGy, **a.** Magnification 1000x (scale: 20  $\mu\text{m}$ ), **b.** Magnification 5000x (scale: 5  $\mu\text{m}$ ).



**Fig.10.** Atomic force microscopy of grafted polystyrene under 30 KGy. (scale:  $1 \times 1 \mu\text{m}$ )

### 3.3. Contact angle analysis

The angles of surface samples of normal polystyrene adjusted by  $\gamma$ -radiated PNIPAAm which were measured at  $10^\circ\text{C}$  and  $37^\circ\text{C}$  temperatures are shown in [Table 1](#). The data for the best-grafted sample (40 KGy pre-irradiated sample) indicate that, at  $10^\circ\text{C}$  and  $37^\circ\text{C}$ , the samples show different contact angles, which is also another reason for the existence of PNIPAAm grafting onto polystyrene surfaces.

**Table 1.** Contact angle for normal and grafted samples

T( $^\circ\text{C}$ )	$\theta_{H2O}$ of Normal samples	$\theta_{H2O}$ of grafted samples
37	94	$61.2 \pm 0.9^\circ$
10	91	$55.3 \pm 1.2^\circ$

The contact angle averages of  $55.3^\circ$  and  $61.2^\circ$  were calculated for  $10^\circ\text{C}$  and  $37^\circ\text{C}$  temperatures. The results indicate a contact angle decrease below  $32^\circ\text{C}$  ( $10^\circ\text{C}$ ), and show the hydrophilic surface feature. The contact angle increases above  $32^\circ\text{C}$  temperature ( $37^\circ\text{C}$ ) which also shows the hydrophobic surface feature.

### 3.4. Differential scanning calorimetry (DSC)

The samples were investigated by thermal analysis using a DSC device (NETZSCHDSC200F3) with a heating rate of 5 degrees per minute from  $0^\circ\text{C}$  to  $60^\circ\text{C}$  in a nitrogen gas atmosphere. A review of the grafted samples' DSC analysis indicates the critical temperature for the grafted PNIPAAm. Figure 11 shows the DSC thermogram from which the curve slope at  $36^\circ\text{C}$  and  $30^\circ\text{C}$  and the critical temperature point at  $32^\circ\text{C}$  are obtained. This shows no significant change in the smart polymer critical temperature during the radiation and grafting process.

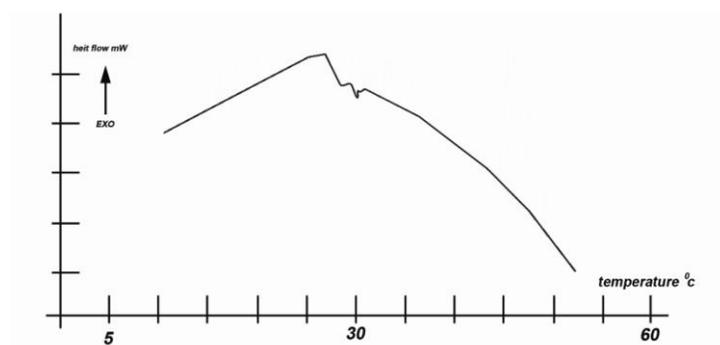


Fig.11. DSC spectra of the grafted polystyrene by gamma ray (radiation dose: 40 KGy)

### 3.5. Cell culture results

The cell attachment and detachment in the grafted polystyrene petri dish were studied with the fibroblast cells (1929) of mouse tail. Figure 12a shows the growth of the fibroblast cells on the grafted Petri dish. In the images, the grafted sample shows good cell adhesion at  $37^\circ\text{C}$ . In Figure 12b the cells have detached spontaneously (cell sheet) from the grafted sample after the temperature dropped below  $10^\circ\text{C}$ .

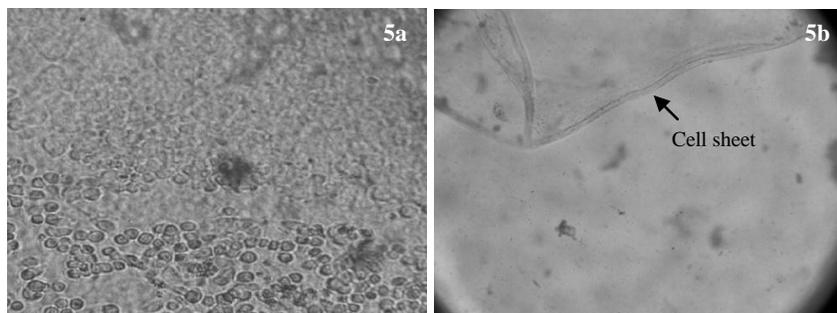


Fig.12. a. The growth of fibroblast cells on the grafted Petri dish at  $37^\circ\text{C}$ , b. the cells detached spontaneously from the grafted sample when the temperature dropped below  $10^\circ\text{C}$ .

#### 4. Conclusion

The effect of a  $\gamma$ -radiation dose on polymer grafting in a polystyrene cell culture dish sample in the air was studied in this article. The FTIR spectrum showed the existence of the grafted polymer on the polystyrene surfaces. The imaging and gravimetric determination of the amount of grafting under radiation dosages of 10, 20, 30, and 40 KGy indicated that an increase in the grafting quantity is achieved by increasing the dose or lengthening the duration of the radiation. Therefore, in this study, optimal grafting was obtained for pre-irradiated samples under a radiation dose of 40 KGy. Increasing the radiation dosage increased the number of sites and free radicals on the polystyrene surface, which led to an increase in the graft. The SEM images showed the morphology of the grafted surfaces at different absorbed dosages; we could clearly observe and compare our graft increases at higher doses. The topology of the surfaces shown in the AFM images also confirmed this claim. The graft thickness of the study samples was about 50-100 nm, which can be attributed equal to the radiation dose intensity to all samples. The contact angles  $55^\circ$  and  $61^\circ$  obtained at  $10^\circ\text{C}$  and  $37^\circ\text{C}$  temperatures, as well as the polymer critical temperature constancy ( $32^\circ\text{C}$ ) measured by the DSC method confirmed that the grafting caused no change in the PNIPAAm operation and function. Thermo-responsive polymers were grafted to dishes covalently, which allowed the fibroblast cells to attach and proliferate at  $37^\circ\text{C}$ . Also cells (cell sheet) detached spontaneously when the temperature decreased below  $32^\circ\text{C}$ , without using enzymes. This characteristic proved that such a type of grafted material has potential as a biomaterial for cell sheet engineering.

#### References

- [1] Park K.D., Suzuki K., Lee WK., Lee JE., Kim YH., Sakurai Y., Okano T., (1996), Platelet adhesion and activation on polyethylene glycol modified polyurethane surfaces. Measurement of cytoplasmic calcium. *ASAIO J.* 42(5):876-881.
- [2] Kikuchi A., Okano T., (2005), Nanostructured designs of biomedical materials applications of cell sheet engineering to functional regenerative tissues and organs. *Journal of Controlled Release.* 101(1-3):69-84.
- [3] Ricardo M.P., da Silva Joao F., Manoand Rui L., (2007), Smart thermo-responsive coatings and surfaces for tissue engineering: switching cell-material boundaries. *Trends in biotechnology.* 25(12):577-583.
- [4] Akiyama Y., Kikuchi A., Yamato M., Okano T., (2004), Ultra thin poly (N-isopropylacrylamide) grafted layer on poly (styrene) surfaces for cell adhesion/detachment control.. *Langmuir.* 20(13):5506-5511.
- [5] Özyürek Z., (2007), Thermo-responsive Glycopolymers via Controlled Radical Polymerization (RAFT) for Biomolecular Recognition. *Eingereicht am: Tag der Verteidigung.* 5:69-72.
- [6] Yamato M., Okano T., (2004), Cell sheet engineering. *Mater Today.* 7:42-47.

- [7] Yanga J., Yamatoa M., Kohnoa C., Nishimotob A., Sekinea H., Fukaib F., Okano T., (2005), cell sheet engineering: Recreating tissues without biodegradable scaffolds, *Biomaterials*. 26(33):6415-6422.
- [8] Youn Mook Lim., Joon Pyo Jeun., Joon Ho Lee., Young Moo Lee., Young Chang Nho., (2007), Cell Sheet Detachment from Poly(N-vinylcaprolactam-co-N-isopropylacrylamide) Grafted onto Tissue Culture Polystyrene Dishes, *J. Ind. Eng. Chem.*, 13(1):21-26.
- [9] Priscila S., Curtia, Márcia R., de Mouraa., Wanderley Veigab., Eduardo Radovanovica., Adley F., Rubiraa, Edvani C. Muniza., (2005), Characterization of PNIPAAm photografted on PET and PS surfaces *Applied Surface Science*. 245:223-233.
- [10] Wang X., (2005), *Responsive Biomaterial Surfaces: Textile Engineering & Biomedical Engineering*.
- [11] Hou Y., Matthews A.R., Ashley M., Smitherman Allen S., (2008), Thermo-responsive nanocomposite hydrogels with cell-releasing behavior. *Biomaterials*. 29:3175-3184.
- [12] Gil E.S., Hudson S.M., (2004), Stimuli-responsive polymers and their bioconjugates. *Progress in Polymer Science*. 29(12):1173-1222.
- [13] Galaev I.Y., Mattiasson B., (2003), Stimulus Responsive Surfaces: Possible Implication for Biochromatography, Cell Detachment and Drug Delivery. *Landes Bioscience*.5(1):68-79.
- [14] Suzuki M., Kishida A., Iwata H., Ikada Y., (1986), Graft copolymerization of acrylamide onto a polyethylene surface pretreated with glow discharge, *macromolecules*. 19 (7):1804-1808.
- [15] Iwata, H., Kishida, A., Suzuki, M., Hata, Y., Ikada, Y., (1988), Oxidation of polyethylene surface by corona, *j polymer. Sci*. 26: 3309-3322.
- [16] Suzuki M., Tamada Y., Iwata H., Ikada Y., (1983), polymer surface modification to attain blood compatibility of hydrophobic polymer in physicochemical aspects of polymer surface. *mittal. k.l. Ed. plenum. New York*. 2:923-941.
- [17] Uyama Y., Ikada Y., (1988), Graft polymerization of acrylamide onto UV-irradiated films, *j.appl.polym.Sci*. 36:1087-1096.
- [18] Fujimoto, K., Takebayashi, Y., Ikada, Y., (1994), Ozone-induced graft polymerization with organophosphate polymer, *j.biomed.mater*.14:13-18.
- [19] Palacios O., Aliev R., Burillo G., (2003), Radiation graft copolymerization of acrylic acid and N-isopropylacrylamide from binary mixtures onto polytetrafluoroethylene. *j.polymer bulletin. springer – verlag*.51:191-197.
- [20] Akerman S., Viinikka P., Svarfvar B., Putkonen K., Jarvinen K., Kontturi K., Nasmann J., Urtti A., Paronen P., 1998, Adsorption of metal cations from aqueous solutions onto the pH responsive poly(vinylidene fluoride grafted poly(acrylic acid) (PVDF-PAA) membrane, *Int J Pharm*.17(1):71-76.
- [21] Gupta B., Mishra S., Saxena S., (2008), Preparation of thermosensitive membranes by radiation grafting of acrylic acid/N-isopropyl acrylamide binary mixture on PET fabric *Radiation Physics and Chemistry*. 77: 553-560.